

Review

Proximal tubule dysfunction in renal diseases - diagnostic significance of proteomics and biomarkers

Disfuncția tubulară proximală în boala renală - importanța diagnostică a proteomicii și biomarkerilor

Ligia Petrica¹, A. Gluhovschi^{2*}, Cristina Gluhovschi¹, Florica Gadalean¹,
C. Balgradean¹, Cristina Groza¹, Silvia Velciov¹

1. Dept. of Nephrology, "Victor Babes" University of Medicine and Pharmacy, County Emergency Hospital Timisoara, Romania

2. Dept. of Obstetrics and Gynecology, "Victor Babes" University of Medicine and Pharmacy, County Emergency Hospital Timisoara, Romania

Abstract

At present time, clinical practice requires advanced studies of plasma and urinary proteomics which should allow the identification, selection, and implementation of plasma and urinary biomarkers useful in the diagnosis of acute and chronic renal diseases, as well as of drug-induced nephrotoxicity. The importance of these biomarkers also resides in their prognostic value and the possibility of their repeated and dynamic assessment, in order to detect early the progression of an acute renal disease to chronic kidney disease, and of the later one to its final stages. In the present paper urinary biomarkers and proteomics, and their application in the accurate diagnosis of tubular lesions in various settings, (such as early diabetic nephropathy, acute kidney injury, glomerular nephropathies, polycystic kidney disease, renal transplantation, Balkan endemic nephropathy, and preeclampsia), are discussed in view of the clinical expertise of the authors in this field.

Keywords: biomarkers, proteomics, tubular lesions

Rezumat

În prezent, practica clinică necesită studii complexe de proteomică plasmatică și urinară, care să permită identificarea, selectarea și implementarea de biomarkeri plasmatici și urinari, utili în diagnosticul bolilor renale acute și cronice, precum și a nefrotoxicității medicamentoase. Importanța acestor biomarkeri rezidă și în valoarea lor prognostică, dar și în posibilitatea unor determinări repetitive, în dinamică, cu scopul de a detecta precoce progresia unei boli renale acute spre cronicizare și a unei boli renale cronice spre stadiile finale. În articolul de față se prezintă biomarkerii și elementele de proteomică urinară cu aplicare în cadrul unui diagnostic de acuratețe al leziunilor tubulare în variate condiții clinice, precum nefropatia diabetică incipientă, leziunea re-

*Corresponding author: Dr. Gluhovschi Adrian, Str.1 Decembrie Nr. 36, Ap.4 300125 Timisoara, Romania,
E-mail: adigluhovschi@yahoo.com

nală acută, nefropatiile glomerulare, rinichiul polichistic, transplantul renal, nefropatia endemic balcanică și preeclampsia). Aceste aspecte sunt discutate în raport cu experiența în domeniu a autorilor.

Cuvinte cheie: biomarkeri, proteomică, leziune tubulară

Urinary proteomics in renal diseases

The term *proteom* implies the total of proteins present in different biological compartments. These compartments may be represented by the whole organism, by a certain tissue, cell or cellular organelle, as well as by biological fluids, such as urine (1). *Proteomics*, is a technology which tries to define the proteic structures contained in these compartments, aims mainly at identifying biomarkers which may be useful in the diagnosis, prognosis and monitoring of different diseases, including renal diseases.

Proteomic studies may be classified as comparative studies (which try to establish the qualitative and quantitative differences between the proteins of the explored compartments), and as descriptive studies (which aim at identifying proteins separated in distinct fractions) (1-3).

Urinary proteins result from plasma filtration, vascular lesions, alterations of the tubular metabolism, and from reabsorption and/or secretion of proteins by cells of the urinary tract. The analysis of proteins is performed either from the urine, or from the urinary exosomes (intracellular vesicles released by the renal epithelial cells as a consequence of intracellular multivesicular bodies fusion with the apical plasma membrane (1).

Urinary proteomics may be used for the accurate diagnosis of tubular lesions in various settings, such as *early diabetic nephropathy*, *acute kidney injury*, *glomerular nephropathies*, *polycystic kidney disease*, *renal transplantation*, *Balkan endemic nephropathy*, and *preeclampsia* (entities which will be discussed in detail in part 2 of the article).

A proteomic analysis of urinary proteins performed in type 2 diabetes mellitus patients with normo-, micro-, and macroalbuminuria showed that several types of urinary proteins were up-regulated, while others were down-regulated.

Variations in the upregulation and downregulation of these proteins were highly predictive of diabetic chronic kidney disease (CKD) progression (4).

The utility of urinary biomarkers identification in the course of both type 1 and type 2 diabetes mellitus derives from the variability in time of albuminuria, from normo- to micro- and macroalbuminuria and reversely, from micro- to normoalbuminuria. Also, highly significant is the stage of albuminuria (5) or even of normoalbuminuria (6-9).

In this context, there can be defined urinary biomarkers of prediction, urinary biomarkers of detection of spontaneous regression from micro- to normoalbuminuria, biomarkers of regression under specific therapy, and biomarkers of progression of diabetic nephropathy and renal function decline (10).

In patients with *acute kidney injury* (AKI), proteomic studies of urinary proteins have revealed various proteins and peptide fragments whose presence in the urine of these patients may define the real onset of the renal lesion. Also, urinary proteomics allows the differentiation of early AKI from established AKI (11).

Urinary proteomics deserves special attention in the diagnosis of *preeclampsia*. A multitude of differential proteins have provided a clue for investigating the mechanism of proteinuria development in preeclampsia. Low urinary angiotensinogen levels were useful for identifying preeclampsia (12,13). Among the biomarkers studied, fibrinogen alpha chain, collagen alpha chain, and uromodulin fragments are of some interest. The markers appear to predict preeclampsia at gestational week 28 with good confidence but not reliably at earlier time points (weeks 12-16 and 20). After prospective validation in other cohorts, these markers may contribute to better prediction, monitoring, and accurate diagnosis of preeclampsia (14).

Urinary biomarkers in the diagnosis of proximal tubule dysfunction

Biological markers, known under the terminology of biomarkers, allow the identification of normal and abnormal processes which occur in various clinical settings, including renal diseases. Biomarkers can be used for the pre-disposition evaluation towards a certain disease, for detection of biological abnormalities, early diagnosis of renal diseases, assessment of prognosis and progression of a disease, and in assessing the response to treatment protocols.

It should be underlined that biomarkers are not involved in the pathogenesis of diseases, being used only as diagnostic and prognostic markers. Recent proteomic studies have identified new urinary biomarkers that include intact or fragmented proteins, which selectively increase or decrease in the course of renal diseases; patterns of proteins which represent specific indicators for various renal diseases; patterns of proteins which have a predictive value for the progression of acute and chronic renal diseases (15,16).

Chronic kidney disease

Diabetic nephropathy

The assessment of proximal tubule dysfunction in the course of diabetes mellitus comprises a panel of urinary biomarkers which allow establishing the early diagnosis of diabetic CKD, even in the stage of normoalbuminuria. The increase in the level of these biomarkers has a predictive value for the occurrence of early diabetic nephropathy.

According to the tubular theory regarding the mechanism of albuminuria in the course of diabetes mellitus and the fact that proximal tubule dysfunction is present in normoalbuminuric patients and precedes the occurrence of albuminuria, urinary biomarkers of proximal tubule dysfunction yield a substantial benefit in the diagnosis and progress of diabetic nephropathy. A vast number of experimental and clin-

ical studies associate the biomarkers of proximal tubule dysfunction with normoalbuminuria, as well as with their utility in the follow-up of albuminuria variability from normo- to microalbuminuria, as well as the possibility to revert micro- into normoalbuminuria (5,17).

Urinary alpha₁-microglobulin is a marker of early proximal tubule dysfunction utilized in various renal diseases, including diabetic nephropathy. This is a protein with low- molecular weight detected in blood and urine as an unbound form, and as a bound form with IgA and albumin. Alpha₁-microglobulin is filtered freely by the glomeruli and reabsorbed by the proximal tubule (5). Increases in the levels of urinary alpha₁-microglobulin has been signaled in normoalbuminuric patients with type 1 diabetes mellitus (18,19) and type 2 diabetes mellitus (8,20,21).

It should be underlined that the increase in urinary alpha₁-microglobulin precedes the occurrence of normoalbuminuria in both type 1 and type 2 diabetes mellitus, a fact which supports the defining intervention of the proximal tubule in glomerular filtered albumin processing. Thus, although renal lesions may be present at the glomerular level before the tubulointerstitial lesions, microalbuminuria does not become detectable as long as the proximal tubule maintains its structural and functional integrity. It is well established by now that the role of the proximal tubule in glomerular filtered albumin processing is fundamental, in the frames of the retrieval and degradation pathways of albumin at the level of the proximal tubule (8,19-21).

Urinary beta₂-microglobulin is a low-molecular weight protein filtered freely at glomerular level and reabsorbed almost entirely by the proximal tubule. This is utilized as an indicator of proximal tubule dysfunction in various renal diseases, including diabetic nephropathy (5). Increases in the levels of urinary beta₂-microglobulin may occur in normoalbuminuric patients with type 2 diabetes mellitus (21-23), thus proving that proximal tubule dysfunction is dissociated from glomerular endothelial dysfunction (8).

In the clinical practice, the correlations between urinary α_1 -microglobulin and urinary β_2 -microglobulin are indicative of proximal tubule dysfunction in early diabetic nephropathy, before the stage of microalbuminuria (8,21,24).

N-acetyl-beta-D-glucosaminidase (NAG) is a lysosomal enzyme of 140 kDa molecular weight localized in the brush border of the proximal tubule cell, and released in the urine as a consequence of proximal tubule lesions (5,25). The levels of NAG also increase in proximal tubule dysfunction evidenced in normoalbuminuric patients with type 1 diabetes mellitus (19,26). In a recent study performed in patients with type 1 diabetes mellitus it has been demonstrated that the transition from micro- to normoalbuminuria is associated with decreased urinary elimination of NAG. This fact supports the hypothesis according to which the earliest renal lesion in type 1 diabetes mellitus is at tubular level and not at glomerular level (27).

Neutrophile gelatinase-associated lipocalin (NGAL) is a lysosomal enzyme localized in the proximal tubule, of 25 kDa molecular weight, and resistant to the activity of proteases. NGAL levels in the urine and plasma increase in the course of acute kidney injury, renal lesions induced by ischaemia/reperfusion, nephrotoxicity, and CKD. Recently, NGAL proved reliable in the diagnosis of early diabetic nephropathy (18). Urinary NGAL levels correlate with proximal tubule dysfunction and renal function in type 2 diabetes mellitus, and have a predictive value for the transition from normo- to microalbuminuria and the occurrence of early diabetic nephropathy (28). Urinary NGAL may be a promising marker for monitoring renal impairment in short-term type 2 diabetes mellitus, thus showing that tubular damage is common in this category of patients (29).

Kidney injury molecule-1 (KIM-1) is a membrane type 1 glycoprotein which contains, in its extracellular domain, immunoglobulin- and mucin-like dimers, with N- and O-glycosylated sites. KIM-1 is expressed at the apical pole of the mem-

brane of the proximal tubular cells, while its ecto-domain is clived and released in the tubular lumen for urinary excretion. KIM-1 is not detected in the normal urine, but it is expressed in large amounts in proximal tubule lesions related to various renal diseases as to the effects of several toxins (5,25).

Increased levels of KIM-1 have also been demonstrated in patients with type 1 diabetes mellitus, in whom the transition from microalbuminuria to normoalbuminuria was accompanied by the decrease in urinary KIM-1 expression. This observation constitutes an argument which favors proximal tubule dysfunction as a preceding event, before glomerular lesions in the course of type 1 diabetes mellitus (27).

Higher urinary tubular damage markers, such as NGAL and KIM-1, were found in type 2 diabetes mellitus patients with glomerular hyperfiltration, probably a direct proof that glomerular hyperfiltration is a deleterious factor for diabetic nephropathy (29).

Liver-type fatty acid-binding protein (L-FABP) is a urinary marker of proximal tubule lesion, its increased levels being detected in all patients with type 1 diabetes mellitus, even in patients without elements of glomerular lesions or albuminuria. Variations in the levels of L-FABP have a predictive value for the progression from normo- to microalbuminuric in patients with type 1 diabetes mellitus (30). Other urinary biomarkers utilized in the diagnosis of proximal tubule dysfunction in type 2 diabetes mellitus are ***fractional excretion of magnesium*** (31), and ***α -glutathion-S-transferase*** (32).

More recently, it has been forwarded the hypothesis according to which ***albuminuria*** per se may be considered as a nephrotoxin and a proximal tubule lesion biomarker. These data are maintained by studies which try to define potential toxins with direct action upon the proximal tubule (Predictive Safety Testing Consortium, US Food and Drug Administration, European Medicines Agency) (33).

Starting from the observation that normal glomeruli filter high amounts of albumin which is

reabsorbed and processed by the proximal tubule, albuminuria occurs when there is an underlying proximal tubule dysfunction (34,35), which may also be induced by albuminuria or peptide fragments derived from the glomerular filtered albumin. The peptides which derive from advanced glycation end-products (AGE_s), such as glycated albumin, might have a potential nephrotoxic effect on the proximal tubule, thus contributing to the occurrence of proximal tubule dysfunction in the course of diabetes mellitus (26). The same nephrotoxic potential may display yet incompletely identified peptides (5).

Glomerular nephropathies

Urinary biomarkers have proved their utility in monitoring primary and secondary chronic glomerulonephritides. Tubulointerstitial lesions are significant in the course of chronic glomerular nephropathies and are related to the degree of renal function decline. Tubular dysfunction may be evidenced by increased eliminations of *leucine-aminopeptidase* and *lactic dehydrogenase* in patients with acute oligoanuric glomerulonephritis. These tubular enzymes have revealed the important contribution of tubular lesions in the occurrence of acute renal failure in the course of acute oligoanuric glomerulonephritis (36).

Some toxic effects of proteinuria on tubular epithelial cells have been considered to be responsible for the tubulointerstitial damage that is frequently associated with glomerular lesions in primary glomerulonephritides. The amount of proteinuria and the degree of tubulointerstitial involvement appeared to be better predictors of functional outcome. A correlation between tubular lesions and renal function has been found, referred to some toxicity of proteinuria on tubular cells. There is a significant relationship between the selectivity of proteinuria and the degree of tubulointerstitial damage. Furthermore, the type of peptide fragments eliminated in the urine initiates toxic effects upon the proximal tubule epithelial cells (37).

In patients with *IgA nephropathy*, fractional excretion of IgG has normalized proportion-

ally to the percentage of glomeruli with segmental sclerosis. Also, the levels of urinary *NGAL*, *NAG*, and *Il-6* increase during active disease (38).

In *membranous nephropathy* there have been described increased urinary eliminations of *NGAL*, *L-FABP*, *NAG*, β_2 -microglobulin, and *IgG*. In *focal and segmental glomerulosclerosis* increase the urinary eliminations of *retinol-binding protein*, while in *minimal change disease* increase the levels of *NAG* and urinary *TGF beta* (38).

In *lupus nephropathy*, the levels of urinary *NGAL*, *NAG*, *TGF β* , *VEGF* and *VCAM-1* correlate with the pathological class of lupus nephropathy (39). *FOXP₃* is a regulator of the development and functionality of T cells. The levels of mRNA *FOXP3* in the urine are elevated in patients with lupus nephropathy, as compared to patients with systemic lupus erythematosus without renal involvement (40). *TWEAK* (tumor necrosis factor-like weak inducer of apoptosis) is a pro-inflammatory cytokine which increases in the urine of patients with lupus nephropathy (41,42).

In *ANCA-mediated vasculitides*, increased urinary excretion of *IgM* was associated with renal function decline (38).

Tubulointerstitial nephropathies

Tubulointerstitial lesions are associated to upper urinary tract infections, to acute and chronic pyelonephritis, respectively. Tubular lesions have been evidenced by assessing urinary eliminations of several tubular enzymes. Thus, increased levels of *NAG* were found as a consequence of tubular cells involvement in tubulointerstitial nephropathies. The assessment of urinary *NAG* allows an accurate follow-up of the response to antibiotherapy of patients with acute and chronic pyelonephritis (43).

Sherman *et al* have revealed increases in urinary *NAG* and β_2 -microglobulin eliminations in patients with urinary tract infections (44). Due to the fact that the urinary levels of *NAG* increase in relation to tubular lesions, this enzyme increases only in upper urinary tract infections, but not in lower urinary tract infections. Thus, the eliminations of *NAG* make the

difference between upper and lower urinary tract infections, respectively (45).

In patients with calcium oxalate renal lithiasis, increased eliminations of tubular enzymes, such as alkaline phosphatase, gamma glutamyl-transpeptidase, lactic dehydrogenase, and NAG may be used in the diagnosis of the patients prone to developing renal calculi (46).

Renal colic is associated with tubular lesions. Consequently, urinary eliminations of tubular enzymes may be elevated, such as is the case of urinary NAG (47). Tubular lesions may occur due to increased urinary tubular enzymes (48).

According to the observations forwarded by Laterza *et al*, the increase in the pressure of the proximal tubule is produced by the passage of stones, a fact which leads to tubular lesions and subsequent increase in urinary eliminations of tubular enzymes (49). Extracorporeal shock-wave lithotripsy is associated with renal lesions, which may be assessed by quantification of urinary enzymes (50).

Proximal tubule dysfunction may occur early in patients with a congenital or surgically acquired single kidney. Tubular lesions are evidenced by increased eliminations of urinary NAG, α_1 -microglobulin, and albuminuria, the latter being now considered a marker of proximal tubule dysfunction, rather than a marker of glomerular lesion (51).

Balkan endemic nephropathy

Balkan endemic nephropathy (BEN) represents a chronic tubulointerstitial nephropathy described amongst renal diseases found in well delimited areas of Serbia, Bulgaria, and Romania. This disease has a slow and progressive evolution towards chronic renal failure, and finally to end-stage renal disease. Due to the fact that its pathogenesis is incompletely elucidated, an early diagnosis of this disease is very difficult to achieve.

Tubulointerstitial lesions are very important in the course of BEN. Thus, tubular lesions have been evidenced by increased urinary eliminations of NAG, leucine-amino-peptidase, lactic dehydrogenase, and lysosim (52).

Acute kidney injury

Acute kidney injury (AKI), defined as an abrupt decrease in renal function and/or of urinary output, may be induced by various causes, such as infections, toxic and ischaemic mechanisms, hypertension, hereditary and metabolic disorders, autoimmune diseases, and acute rejection of the renal allograft (53).

The incidence of acute kidney injury is increasing to epidemic proportions. Development of AKI leads to excessive morbidity and mortality, prolonged hospitalization, and increased healthcare costs.

Although serum creatinine is typically used for diagnosis of AKI, it is an insensitive and unreliable biomarker during acute changes in kidney function. The serum creatinine levels do not increase until about half of the kidney function is lost (54,55).

Early detection of AKI requires more reliable and specific biomarkers in order to accurately diagnose acute tubular lesions. Emerging and already established biomarkers are widely used in the early diagnosis of AKI, especially in critically ill patients in the intensive care unit (ICU) (54,56).

The biomarkers indicative of AKI are classified as ***functional markers (enzymatic markers:*** NAG, α/π GST, γ -glutamyltranspeptidase; ***inflammatory markers:*** IL-18, NGAL), and ***structural markers*** (KIM-1, Na/H-exchanger isoform 3, L-FABP). These markers are eliminated in high proportions in the urine of patients with AKI due to tubular lesions.

Furthermore, biomarkers of AKI may be represented by ***low-molecular weight proteins***, normally filtered by the glomeruli and/or metabolised by the normal tubular epithelial cells (cystatin C, α_1 -microglobulin, β_2 -microglobulin, retinol-binding protein) (57). These biomarkers are useful in differentiating patients with ***established acute renal lesions*** from patients hospitalized for non-renal diseases, chronic kidney disease, and from healthy control subjects (57).

In the emergency departments and in the ICU, several biomarkers, such as NGAL and

NHE3, allow differentiating of cases with established acute renal lesions from increases in serum creatinine due to prerenal azotaemia (58,59).

In clinical practice, early detection of acute renal lesions, within hours of the onset, permits identification of the therapeutic window, in which adequate treatment strategies may improve outcomes, before established lesions and functional modifications occur (57).

Cystatin C has a very high sensitivity in the early detection of renal function modifications, even in mild renal function impairment (60). This biomarker has a diagnostic value superior to serum creatinine in the evaluation of AKI due to its short half-time (61).

Urinary Neutrophil gelatinase-associated lipocalin (NGAL) increases significantly in the proximal tubules and in the first voided urine after AKI through ischaemic mechanisms (62,63). NGAL occurs in the urine before other urinary biomarkers of AKI, relevant for proximal tubule dysfunction, such as NAG and β_2 -microglobulin. Therefore, NGAL has been named "renal troponin". Similar aspects have been described in AKI through nephrotoxic mechanisms (64).

The increased levels of urinary NGAL allow differentiating intrinsic AKI from prerenal azotaemia, and also these may serve in stratification of patients with AKI according to the RIFLE criteria (65).

Kidney injury molecule-1 (KIM-1) represents a rapid, sensitive, non-invasive, and reproducible biomarker, used for the early detection of AKI induced by cisplatin and through ischaemic mechanisms (63,66).

N-acetyl-beta-D-glucosaminidase (NAG). Increased levels of urinary NAG have been found in tubulointerstitial nephropathies, AKI, drug-induced nephrotoxicity, diabetic nephropathy, and acute renal allograft rejection (63,67).

Liver-type fatty acid-binding protein (L-FABP) increases in the urine before serum BUN and creatinine, in cisplatin and ischaemic mechanisms induced AKI (68). L-FABP also increases in patients with severe sepsis (16).

Exosomal transcription factors may be utilized in the identification of acute renal lesions. Exosomes are microvesicles excreted by normal and injured renal epithelial cells. Urinary exosomes may be released by all segments of the nephron, including podocytes. Exosomes are vesicles of 50-90 nm, created within a cell when a segment of the cell membrane is invaginated and undergoes a process of endocytosis. The internalized membrane segment is fragmented in small vesicles, which may be externalized. Detection of exosomes which contain activator transcription factor 3 (ATF₃) in the urine is indicative of AKI before a rise in serum creatinine (69). These exosomes contain transcription factors which may be activated by various stimuli and may be detected in the urine of patients with AKI (69), and with chronic kidney disease (15,63).

L1 cell adhesion molecule (CD 171) is a membrane glycoprotein which belongs to the immunoglobulin superfamily. The levels of this glycoprotein increase in the urine of patients with acute tubular necrosis, as compared with patients with prerenal azotaemia (16).

Netrins are laminin-like molecules, with a distinctly organised domain. Netrin-1 increases before serum creatinine in patients with AKI induced through ischaemic mechanisms, cisplatin, folic acid, and endotoxins (16).

Aprotinin, also known as a pancreatic trypsin inhibitor, is a protein which increases in the urine of patients with AKI after cardio-pulmonary by-pass (70).

Nephronectin (NPNT), ligand of $\alpha_8\beta_1$ integrin, is expressed by the urethelial bud during renal morphogenesis. The levels of urinary nephronectin increase in AKI through nephrotoxic mechanisms, reaching its maximal levels in the course of tubular cells regeneration in the onset stage of AKI, and in the recovery period after AKI (71).

Other urinary biomarkers released by the injured proximal tubule cells are **glutathion-S-transferase**, **alkaline phosphatase**, and **γ glutamyl-transpeptidase**, which are indicative of AKI induced by nephrotoxic mechanisms, and of

diabetic nephropathy (16). Also, urinary alpha₁-microglobulin and beta₂-microglobulin may serve in the diagnosis of AKI (16).

Leucin-aminopeptidase increases in the urine of patients with AKI after exposure to mercury and after iodinated contrast media, such as NN'-diacetyl-3,5diamino-2,4,6-triiodobenzoic acid, utilized for i.v. urography or aortic angiography (72).

Also, drug-induced nephrotoxicity has been evidenced by increased urinary eliminations of NAG (cysplatin) (73), rifampicin (74), aspirin (75), gentamicin (76), and amikacin (47).

It should be stressed that aminoglycosides accumulate and concentrate in the lysosomes within the proximal tubule cells. Lesions in these cells and the cellular organelle are associated with increased urinary NAG eliminations. Mild renal dysfunction due to tubular lesions may be detected early by assessment of NAG (77).

Preeclampsia

High urinary levels of tubular enzymes have been observed during preeclampsia. Manescu *et al* have shown increased urinary eliminations of leucine-aminopeptidase in patients with preeclampsia (primary toxemia of pregnancy). The highest levels of enzymuria were observed in severe forms of preeclampsia, especially in the complicated cases (superimposed eclampsia, uteroplacental apoplexia). The determination of urinary leucine-aminopeptidase may be a relatively accurate test in estimating severity and course of toxemia of pregnancy (78). The cause of high levels of enzymuria may be related to a spastic factor.

An increased cell permeability in the proximal tubule cells, induced either by hypoxia or by humoral factors, such as an increase in urinary oxytocinase excretion can also be an important factor.

Also, increased eliminations of NAG have been reported during preeclampsia. This increase was much higher than that corresponding to the gestational age. This fact is due to the tissue ischaemia that precedes glomerular endotheliosis, but also tubular lesions caused by arteriolar vasoconstriction may have an important role (79).

Concluding remarks

At present time, clinical practice requires advanced studies of plasma and urinary proteomics which should allow identification, selection, and implementation of plasma and urinary biomarkers, useful in the diagnosis of acute and chronic renal diseases, as well as of drug-induced nephrotoxicity. The importance of these biomarkers also resides in their prognostic value and the possibility of their repeated and dynamic assessment, in order to detect early the progression of an acute renal disease to chronic kidney disease, and of the later one to its final stages.

List of abbreviations

AGES – advanced glycation end-products
 AKI – acute kidney injury
 ANCA – Anti-neutrophil cytoplasmic antibody
 ATF3 – activator transcription factor 3
 BEN – Balkan endemic nephropathy
 BUN – blood urea nitrogen
 CKD – chronic kidney disease
 α/π GST – α/π Glutathione S-transferase
 ICU – Intensive-care unit
 IgA – Immunoglobulin A
 IgG – Immunoglobulin G
 Il-6 – interleukin 6
 Il-18 – interleukin 18
 KIM-1 – Kidney injury molecule-1
 L-FABP – Liver-type fatty acid-binding protein
 NAG – N-acetyl-beta-D-glucosaminidase
 NGAL – Neutrophile gelatinase-associated lipocalin
 NPNT – Nephronectin
 TGF beta – Transforming growth factor beta
 TWEAK – tumor necrosis factor-like weak inducer of apoptosis
 VCAM-1 – Vascular cell adhesion molecule-1
 VEGF – Vascular endothelial growth factor

References

1. Welberry-Smith MP, Banks RE, Wood SL, Lewington AJ, Selby PJ. Application of proteomic analysis to the study of renal diseases. *Nat Rev Nephrol* 2009; 5:701-712.
2. Granier C, Makni K, Molina L, Jardin-Watelet B, Ayadi H, Jarraya F. Gene and protein markers of diabetic nephropathy. *Nephrol Dial Transplant* 2008; 23:792-799.

3. Mischak H, Massy ZA, Jankowski J. Proteomics in uremia and renal disease. *Semin Dial* 2009; 22:409-416
4. Rao PV, Lu X, Standley M, Pattee P, Neelima G, Girish G, et al. Proteomic identification of urinary biomarkers of diabetic nephropathy. *Diabetes Care* 2007; 30:629-637
5. Merchant ML, Klein JB. Urinary Proteomics and Candidate Biomarker Discovery for Diabetic Nephropathy, in Edelstein CL (ed.) *Biomarkers in Kidney Disease*, Academic Press, Elsevier 2011, p.352-368
6. Perkins BA, Ficociello LH, Roshan B, Warram JH, Krolewski AS. In patients with type 1 diabetes and new-onset microalbuminuria the development of advanced chronic kidney disease may not require progression to proteinuria. *Kidney Int* 2010; 77(1):57-64
7. MacIsaac RJ, Tsalamandris C, Panagiotopoulos S, Smith TJ, McNeil KJ, Jerums G. Nonalbuminuric renal insufficiency in type 2 diabetes. *Diabetes Care* 2004; 27:195-200
8. Petrica L, Petrica M, Vlad A, Jianu DC, Gluhovschi G, Ianculescu C, et al. Proximal tubule dysfunction is dissociated from endothelial dysfunction in normoalbuminuric patients with type 2 diabetes mellitus: a cross-sectional study. *Nephron Clin Pract* 2011; 118:c155-c164
9. Petrica L, Vlad A, Petrica M, Jianu CD, Gluhovschi G, Gadalean F, et al. Pioglitazone delays proximal tubule dysfunction and improves cerebral vessels endothelial dysfunction in normoalbuminuric patients with type 2 diabetes mellitus. *Diabetes Res Clin Pract* 2011; 94(1):22-32
10. Ameer RB, Molina L, Bolvin C, Kifagi C, Jarraya F, Ayadi H, et al. Proteomic approaches for discovering biomarkers of diabetic nephropathy. *Nephrol Dial Transplant* 2010; 25:2866-2875
11. Metzger J, Kirsch T, Schiffer E, Ulger P, Menten E, Brand K, et al. Urinary excretion of twenty peptides forms an early and accurate diagnostic pattern of acute kidney injury. *Kidney Int* 2010; 78:1252-1262
12. Chen G, Zhang Y, Jin X, Zhang L, Zhou Y, Niu J, et al. Urinary proteomics analysis for renal injury in hypertensive disorders of pregnancy with iTRAQ labeling and LC-MS/MS. *Proteomics Clin Appl* 2011; 5(5-6):300-310
13. Lee SM, Park JS, Norwitz ER, Kim SM, Kim BJ, Park CW, et al. Characterization of discriminatory urinary proteomic biomarkers for severe preeclampsia using SELDI-TOF mass spectrometry. *J Perinat Med* 2011; 39:391-396
14. Carty DM, Siwy J, Brennand JE, Züribig P, Mullen W, Franke J, et al. Urinary proteomics for prediction of preeclampsia. *Hypertension* 2011; 57(3):561-569
15. Tesch GH. Review: Serum and urine biomarkers of kidney disease: A pathophysiological perspective. *Nephrology (Carlton)* 2010; 15:609-616
16. Edelstein CL, Faubel S. Biomarkers in acute kidney injury, in Edelstein CL *Biomarkers in Kidney Disease*. Academic Press, Elsevier 2011, p.179-220
17. Thongboonkerd V. Searching for Novel Biomarkers and New Therapeutic Targets of Diabetic Nephropathy Using Proteomic Approaches in Thongboonkerd V (ed.). *Proteomics in Nephrology - Towards Clinical Applications*, Karger 2008, p.37-53
18. Korpinen E, Teppo AM, Hukkanen L, Akerblom HK, Grönhagen-Riska C, Vaarla C. Urinary transforming growth factor- β 1 and α 1-microglobulin in children and adolescents with type 1 diabetes. *Diabetes Care* 2000; 23:664-668
19. Pfliegerer S, Zimmerhackl LB, Kinne R, Manz F, Schuler G, Brandis M. Renal proximal and distal tubular function is attenuated in diabetes mellitus type 1 as determined by the renal excretion of alpha 1-microglobulin and Tamm-Horsfall protein. *Clin Investig* 1993;71(12):972-977
20. Hong CY, Hughes K, Chia KS, Ng V, Ling SL. Urinary alpha1-microglobulin as a marker of nephropathy in type 2 diabetic Asian subjects in Singapore. *Diabetes Care* 2003; 26:338-342
21. Petrica L, Petrica M, Vlad A, Jianu DC, Gluhovschi Gh, Ianculescu C, et al. Nephro- and neuroprotective effects of rosiglitazone versus glimepiride in normoalbuminuric patients with type 2 diabetes mellitus: a randomized controlled trial. *Wien Klin Wochenschr* 2009; 121:765-775
22. Aksun SA, Ozmen D, Ozmen B, Parildar Z, Mutaf I, Turgan N, et al. Beta2-microglobulin and cystatin C in type 2 diabetes: assessment of diabetic nephropathy. *Exp Clin Endocrinol Diabetes* 2004; 112:195-200
23. Kalansooriya A, Holbrook I, Jennings P, Whiting PH. Serum cystatin C, enzymuria, tubular proteinuria and early renal insult in type 2 diabetes. *Br J Biomed Sci* 2007; 64:121-123
24. Holmquist P, Torffvit O. Tubular function in diabetic children assessed by Tamm-Horsfall protein and glutathione S-transferase. *Pediatr Nephrol* 2008; 23(7):1079-1083
25. Vaidya VS, Ford GM, Waikar SS, Wang Y, Clement MB, Ramirez V, et al. A rapid urine test for early detection of kidney injury. *Kidney Int* 2009; 76:108-114
26. Kern EF, Erhard P, Sun W, Genuth S, Weiss MF. Early urinary markers of diabetic kidney disease: a nested case-control study from the Diabetes Control and Complications Trial (DCCT). *Am J Kidney Dis* 2010; 55(5):824-834
27. Vaidya VS, Niewczas MA, Ficociello LH, Johnson AC, Collings FB, Warram JH, et al. Regression of microalbuminuria in type 1 diabetes is associated with lower levels of urinary tubular injury biomarkers, kidney injury molecule-1, and N-acetyl- β -D-glucosaminidase. *Kidney Int* 2011; 79(4):464-470
28. Yang YH, He XJ, Chen SR, Wang L, Li EM, Xu LY. Changes of serum and urine neutrophil gelatinase-associated lipocalin in type-2 diabetic patients with nephropathy: one year observational follow-up study. *Endocrine* 2009; 36(1):45-51

29. Fu WJ, Xiong SL, Fang YG, Wen S, Chen ML, Deng RT, et al. Urinary tubular biomarkers in short-term type 2 diabetes mellitus patients: a cross-sectional study. *Endocrine* 2012; 41(1):82-88
30. Nielsen SE, Sugaya T, Hovind P, Baba T, Parving HH, Rossing P. Urinary liver-type fatty acid-binding protein predicts progression to nephropathy in type 1 diabetic patients. *Diabetes Care* 2010; 33(6):1320-1324
31. Futrakul N, Vongthavarat V, Sirisalipotch S, Chairatanarat T, Futrakul P, Suwanwalaikorn S. Tubular dysfunction and hemodynamic alteration in normoalbuminuric type 2 diabetes. *Clin Hemorheol Microcirc* 2005; 32(1):59-65
32. Cawood TJ, Bashir M, Brady J, Murray B, Murray PT, O'Shea D. Urinary collagen IV and π GST: potential biomarkers for detecting localized kidney injury in diabetes--a pilot study. *Am J Nephrol* 2010; 32(3):219-225
33. Yu JY, An XF, Liu JS, Ten SC, Wang X, Zhao Y, et al. Plasma sRAGE is not associated with urinary microalbumin excretion in type 2 diabetic nephropathy at the early stage. *Diabetes Res Clin Pract* 2010; 87(2):157-160
34. Russo LM, Sandoval RM, McKee M, Osicka TM, Collins AB, Brown D, et al. The normal kidney filters nephrotic levels of albumin retrieved by proximal tubule cells: retrieval is disrupted in nephrotic states. *Kidney Int* 2007; 71(6):504-513
35. Russo LM, Sandoval RM, Campos SB, Molitoris BA, Comper WD, Brown D. Impaired tubular uptake explains albuminuria in early diabetic nephropathy. *J Am Soc Nephrol* 2009; 20(3):489-494
36. Zosin C, Mănescu N, Gluhovschi G, Schwarzkopf A, Golea O. Enzymatic changes in the urine (L.A.P. and L.D.H.) in acute oligoanuric glomerulonephritis. *Med Welt* 1973; 24(20):813-816
37. Bazzi C, Petrini C, Rizza V, Arrigo G, D'Amico G. A modern approach to selectivity of proteinuria and tubulointerstitial damage in nephrotic syndrome. *Kidney Int* 2000; 58(4):1732-1741
38. Arthur JM, Budisavljevic MN, Janech MG. Biomarkers in Glomerular Disease, in Edelstein CL. Biomarkers in Kidney Disease. Academic Press, Elsevier 2011, p.368-383
39. Avihingsanon Y, Phumesin P, Benjachat T, Akkasilpa S, Kittikowit V, Praditpornsilpa K, et al. Measurement of urinary chemokine and growth factor messenger RNAs: a noninvasive monitoring in lupus nephritis. *Kidney Int* 2006; 69:747-753
40. Nakayamada S, Saito K, Nakano K, Tanaka Y. Activation signal transduction by beta1 integrin in T cells from patients with systemic lupus erythematosus. *Arthritis Rheum* 2007; 56:1559-1568
41. Schwartz N, Rubinstein T, Burkly LC, Collins CE, Blanco I, Su L, et al. Urinary TWEAK as a biomarker of lupus nephritis: a multicenter cohort study. *Arthritis Res Ther* 2009; 11:R143
42. Pitashny M, Schwartz N, Qing X, Hojaili B, Aranow C, Mackay M, et al. Urinary lipocalin-2 is associated with renal disease activity in human lupus nephritis. *Arthritis Rheum* 2007; 56:1894-1903
43. Gluhovschi G, Velciov S, Kaycsa A, Trandafirescu V, Schiller A, Petrica L, et al. Bestimmung von N-Azetyl-b-D-Glukosaminidase (NAG) bei tubulären Läsionen während einer Nierenkolik mit und ohne assoziierte Harnwegsinfektionen (HWI). *Nieren- und Hochdruckkrankheiten* 2005; 34(10):458-463
44. Sherman RL, Drayer DE, Leyland-Jones BR, Reidenberg MM. N-acetyl-beta-glucosaminidase and beta 2-microglobulin. Their urinary excretion in patients with renal parenchymal disease. *Arch Intern Med* 1983; 143(6):1183-1185
45. Mengoli C, Lechi A, Arosio E, Rizzotti P, Lechi C, Corgnati A, et al. Contribution of four markers of tubular proteinuria in detecting upper urinary tract infections. A multivariate analysis. *Nephron* 1982; 32(3):234-238
46. Prinsen JH, Günther H, Breuer J. Determination of enzyme activities in urine of patients with calcium oxalate calculi. *J Clin Chem Clin Biochem* 1986; 24(12):1001-1007
47. Gluhovschi G, Velciov S, Kaycsa A, Trandafirescu V, Schiller A, Petrica L, et al. Die Verwendung der urinären NAG-Ausscheidung für die Bewertung der Entwicklung und Nephrotoxizität der HWI-Behandlung mit Amikacin. *Nieren- und Hochdruckkrankheiten* 2004; 33(6):285-288
48. Khan SR, Shevock PN, Hackett RL. Urinary enzymes and calcium oxalate urolithiasis. *J Urol* 1989; 142(3):846-849
49. Laterza G, Santini P, Russo G, Scibinetti F, Gentile V, Liberti M, et al. Partial and multiple correlations between 4 types of enzymuria in 2 groups of chronic nephropathic patients. *Minerva Med* 1985; 76(3-4):99-104
50. Jung K, Mattenheimer H, Burchardt U. Urinary Enzymes in Clinical and Experimental Medicine. Springer-Verlag, Berlin 1992; p.169
51. Gadalean FN, Gluhovschi G, Trandafirescu V, Petrica L, Velciov S, Bozdog G, et al. Estimated glomerular filtration rate in patients with surgically acquired single kidney compared with patients with congenital single kidney: implications for kidney transplant from live donors. *Exp Clin Transplant* 2010; 8(3):228-236
52. Gluhovschi G, Sabo I, Zosin C, Manescu M. Biochemical changes in Endemic (Balkan) Nephropathy. Proceedings of the 4th Symposium on endemic (Balkan) nephropathy Nis 1979; 209-214
53. Srisawat N, Hoste EE, Kellum JA. Modern classification of acute kidney injury. *Blood Purif* 2010; 29:300-307
54. Soni SS, Ronco C, Katz N, Cruz DN. Early diagnosis of acute kidney injury: the promise of novel biomarkers. *Blood Purif* 2009; 28(3):165-174
55. Dennen P, Parikh CR. Biomarkers of acute kidney injury: can we replace serum creatinine? *Clin Nephrol*

- 2007; 68(5):269-278
56. Ronco C. N-GAL: diagnosing AKI as soon as possible. *Crit Care* 2007; 11(6):173
57. Siew ED, Ware LB, Izkizler TA. Biological markers of acute kidney injury. *J Am Soc Nephrol* 2011; 22:810-820
58. Nickolas TL, O'Rourke MJ, Yang J, Sise ME, Canetta PA, Barasch N, et al. Sensitivity and specificity of a single emergency department measurement of urinary neutrophil gelatinase-associated lipocalin for diagnosing acute kidney injury. *Ann Intern Med* 2008; 148:810-819
59. du Cheyron D, Daubin C, Poggioli J, Ramakers M, Houillier P, Charbonneau P, et al. Urinary measurement of Na⁺/H⁺ exchanger isoform 3 (NHE3) protein as new marker of tubule injury in critically ill patients with ARF. *Am J Kidney Dis* 2003; 42:497-506
60. Mussap M, Dalla Vestra M, Fioretto P, Saller A, Varagnolo M, Nosadini R, et al. Cystatin C is a more sensitive marker than creatinine for the estimation of GFR in type 2 diabetic patients. *Kidney Int* 2002; 61:1453-1461
61. Herget-Rosenthal S, Marggraf G, Hüsing J, Göring F, Pietruck F, Janssen O, et al. Early detection of acute renal failure by serum cystatin C. *Kidney Int* 2004; 66:1115-1122
62. Mishra J, Ma Q, Prada A, Mitsnefes M, Zahedi K, Yang J, et al. Identification of neutrophil gelatinase-associated lipocalin as a novel early urinary biomarker for ischemic renal injury. *J Am Soc Nephrol* 2003; 14:2534-2543
63. Fassett FG, Venuthurupalli SK, Gobe GC, Coombes JS, Cooper MA, Hoy WE. Biomarkers in chronic kidney disease: a review. *Kidney Int* 2011; 80:806-821
64. Mishra J, Mori K, Ma Q, Kelly C, Barasch J, Devarajan P. Neutrophil gelatinase-associated lipocalin: a novel early urinary biomarker for cisplatin nephrotoxicity. *Am J Nephrol* 2004; 24:307-315
65. Singer E, Elger A, Elitok S, Kettritz R, Nickolas TL, Barasch J, et al. Urinary neutrophil gelatinase-associated lipocalin distinguishes pre-renal from intrinsic renal failure and predicts outcomes. *Kidney Int* 2011; 80:405-414
66. Vaidya VS, Ramirez V, Ichimura T, Bobadilla NA, Bonventre JV. Urinary kidney injury molecule-1: a sensitive quantitative biomarker for early detection of kidney tubular injury. *Am J Physiol Renal Physiol* 2006; 290:F517-F529
67. Han WK, Waikar SS, Johnson A, Betensky RA, Dent CL, Devarajan P, et al. Urinary biomarkers in the early diagnosis of acute kidney injury. *Kidney Int* 2008; 73:863-869
68. Negishi K, Noiri E, Doi K, Maeda-Mamiya R, Sugaya T, Portilla D, et al. Monitoring of urinary L-type fatty acid-binding protein predicts histological severity of acute kidney injury. *Am J Pathol* 2009; 174:1154-1159
69. Zhou H, Cheruvanky A, Hu X, Matsumoto T, Hiramatsu N, Cho ME, et al. Urinary exosomal transcription factors, a new class of biomarkers for renal disease. *Kidney Int* 2008; 74:613-621
70. Wagener G, Gubitosa G, Wang S, Borregaard N, Kim M, Lee HT. Increased incidence of acute kidney injury with aprotinin use during cardiac surgery detected with urinary NGAL. *Am J Nephrol* 2008; 28:576-582
71. Cheng CW, Ka SM, Yang SM, Shui HA, Hung YW, Ho PC, et al. Nephronectin expression in nephrotoxic acute tubular necrosis. *Nephrol Dial Transplant* 2008; 23:101-109
72. Bignon H, Gluhovschi G, Mănescu N, Zosin C. Changes in leucineaminopeptidase activity in the urine caused by contrast media. *Fortschr Geb Rontgenstr Nuklearmed* 1973; 119(3):343-346
73. Litterst C, Smith JH, Smith MA, Uozumi J, Copley M. Sensitivity of urinary enzymes as indicators of renal toxicity of the anticancer drug cis-platin. *Uremia Invest* 1985-1986; 9(2):111-117
74. Kumar BD, Prasad CE, Krishnaswamy K. Detection of rifampicin-induced nephrotoxicity by N-acetyl-3-D-glucosaminidase activity. *J Trop Med Hyg* 1992; 95(6):424-427
75. Lockwood TD, Bosmann HB. The use of urinary N-acetyl-beta-glucosaminidase in human renal toxicology. II. Elevation in human excretion after aspirin and sodium salicylate. *Toxicol Appl Pharmacol* 1979; 49(2):337-345
76. Carver MP, Monteiro-Riviere NA, Brown TT, Riviere JE. Dose-response studies of gentamicin nephrotoxicity in rats with experimental renal dysfunction. II. Polyvinyl alcohol glomerulopathy. *Toxicol Appl Pharmacol* 1985; 80(2):264-273
77. Fogazzi GB, Fenili D. *Urinalysis and microscopy*. Oxford Textbook of Clinical Nephrology 2nd Edition. Oxford University Press 1997; vol.1, p.21
78. Manescu N, Gluhovschi G, Serban V, Ianovici E, Zosin C. Serum and urinary leucine-aminopeptidase in toxemia of pregnancy. *Schweiz Z Gynak Geburtsh* 1978; 3:201-207
79. Pérez-Blanco FJ, Sanabria MC, Huertas JM, Cantero J, Rodríguez-Cuartero A. Urinary N-acetyl-beta-glucosaminidase in the prediction of preeclampsia. *Clin Nephrol* 1998; 50(3):169-171